

## Protective effect of antioxidant medicinal herbs, Rosemary and Parsley, on subacute aflatoxicosis in *Oreochromis niloticus*

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**Abstract:** The object of this study was to conduct the ability of two medicinal herbs, namely rosemary and parsley, for amelioration of aflatoxicosis in *Oreochromis niloticus*. Two herbs' extracts at three concentrations of either (0, 2 and 4 g kg<sup>-1</sup> B.W. divided into 2 doses at the start and the 6<sup>th</sup> day of the experiment) and three concentrations of aflatoxin B<sub>1</sub>, (AFB<sub>1</sub> 0, 9 and 18 mg kg<sup>-1</sup> B.W. as a single intraperitoneal administration) were tested either individually or in combination. The herbs and AFB<sub>1</sub> were dissolved in Dimethylsulphoxide (DMSO 25%) and injected to fish groups. Sixteen groups of fish were investigated in this study, where A group (control) was injected with saline 0.89%, group B injected with DMSO (control solvent), groups F<sub>1</sub> and F<sub>2</sub> were injected with AFB<sub>1</sub> alone (9 and 18 mg kg<sup>-1</sup> B.W. respectively), R<sub>1</sub> and R<sub>2</sub> groups were injected with rosemary alone (2 and 4 g kg<sup>-1</sup> B.W., respectively), groups F<sub>1</sub>R<sub>1</sub>, F<sub>1</sub>R<sub>2</sub>, F<sub>2</sub>R<sub>1</sub> and F<sub>2</sub>R<sub>2</sub> were injected with AFB<sub>1</sub> + rosemary, while groups P<sub>1</sub> and P<sub>2</sub> were injected with parsley alone (2 and 4 g kg<sup>-1</sup> B.W., respectively); however, F<sub>1</sub>P<sub>1</sub>, F<sub>1</sub>P<sub>2</sub>, F<sub>2</sub>P<sub>1</sub> and F<sub>2</sub>P<sub>2</sub> groups were injected with AFB<sub>1</sub> + parsley. At the 12<sup>th</sup> day of the experiment, blood and liver samples were taken from each group. The results indicated that the AFB<sub>1</sub> injected groups revealed a significant increase in mortality rate (MR%) compared with AFB<sub>1</sub>-not injected, group F<sub>2</sub> was the highest while F<sub>1</sub>R<sub>1</sub> and F<sub>1</sub>P<sub>1</sub> were the lowest in MR% among all AFB<sub>1</sub> injected fish groups. Also, AFB<sub>1</sub> led to reduction of haemoglobin (Hb), total protein (TP) and globulin (Gl) concentrations and increase in activity of aspartate aminotransferase (AST) and alanine aminotransferase (ALT). These alterations were significantly ameliorated when fish were injected with herbs' extracts. AFB<sub>1</sub> residues showed that the herbs level of 2g kg<sup>-1</sup> B.W. have higher potency of reducing the AFB<sub>1</sub> residues than the level of 4 g kg<sup>-1</sup> B.W. in case of AFB<sub>1</sub> level 9 mg kg<sup>-1</sup> B.W. While, in case of AFB<sub>1</sub> level 18 mg kg<sup>-1</sup> B.W., the groups F<sub>2</sub> and F<sub>2</sub>P<sub>1</sub> showed absence of AFB<sub>1</sub> residues. Microscopically, AFB<sub>1</sub> presented histopathological changes in hepatopancrease which increased in severity with increasing AFB<sub>1</sub> level. These lesions may become less severer in all fish groups injected with AFB<sub>1</sub> combined with herbs' extracts especially with the lowest levels of herbs' extracts and AFB<sub>1</sub>. So, this study concluded that either of rosemary or parsley was found to be safe and successful in protection from aflatoxicosis, particularly at the low level.

**Key words:** Tilapia – Aflatoxicosis – Parsley – Rosemary- Residues.

### INTRODUCTION

Aflatoxins are secondary metabolites produced by the ubiquitous fungi *Aspergillus flavus* and *A. parasiticus*. Aflatoxin B<sub>1</sub> (AFB<sub>1</sub>) has the highest potency as a toxin and is classified as group 1 carcinogen by International Agency for Research on Cancer (IARC, 1993). Aflatoxin has to be activated in order to exert its carcinogenic effect. Also, the free radical and reactive oxygen species (ROS) may, in part, be responsible for the carcinogenic activity of AFB<sub>1</sub> (Shen *et al.*, 1996). So, inhibition of cytochromes (CYP<sub>450</sub>) and/or stimulation of the antioxidant defence system,  $\alpha$ -tocopherol, ascorbate and reduced glutathione (GSH), may reduce the risk of AFB-mediated carcinogenesis.

Previous studies referred to many medicinal herbs that serve as sources of antioxidant which have antiaflatoxicogenic effects such as *Thonningia sanguinea* (Gyamfi and Ariya, 1998 and Gyamfi *et al.*, 1999), *Cymbopogon citratus* Staf and *Murdannia ioriformis* (Vinitketkumnien *et al.*, 1999), *Oldenlandia diffusa* and *Seutellaria babata* (Wong *et al.*, 1993), as well as *Ocimum sanctum* (Rastogi *et al.*, 2007).

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While, parsley (*Petroselinum crispum*) and rosemary (*Rosmarinus officinalis*) are medicinal herbs which are widely used around the world that have shown a good antioxidant activity (Lampe *et al.*, 2000; Hinneburg *et al.*, 2006; Sacan Ozsoy *et al.*, 2006 and Caillet *et al.*, 2007). As well as, Lampe *et al.* (2000) reported that parsley has been shown to inhibit CYP<sub>1A2</sub> in human and it was suggested that the inhibition was possibly related to its phytochemical content. Natural polyphenols found in rosemary have not only potent antioxidant activities but also anticarcinogenic properties. Rosemary components inhibit both the initiation and tumour promotion stages of carcinogenesis in mouse and rat models (Tokuda *et al.*, 1986; Singletary and Nelshoppen, 1991; Yasukawa *et al.*, 1991 and Huang *et al.*, 1994). Also, Offord *et al.* (1997) reported that rosemary extract strongly inhibits metabolic activation of two important human procarcinogens, AFB<sub>1</sub> and benzo (a) pyrene. Additionally, the medicinal properties of parsley are stimulant, diuretic, carminative, emmenagogue, antipyretic and anti-inflammatory, while the medicinal properties of rosemary are mild irritant, carminative, stimulant and diaphoretic (Peter, 2001). The last author reported that the lethal dose (LD<sub>50</sub>) of rosemary and parsley essential oils determined in rats are > 5 and 1-5g/kg B.W., respectively. The nutritive values of rosemary and parsley were reported by Farrel (1990), it was higher in parsley than in rosemary as shown from the following:

**Nutritive value of rosemary and parsley (approximate composition/ 100g of edible portion)**

Composition	Rosemary	Parsley
Energy (Kcal)	331	276
Protein (g)	4.9	22.4
Fat (g)	15.2	4.4
Total carbohydrates (g)	64.1	51.7
Fibre (g)	17.7	10.3
Ash (g)	6.5	12.5
Ca (mg)	1280	1468
Fe (mg)	29	98
Mg (mg)	220	249
P (mg)	70	351
K (mg)	955	3805
Na (mg)	50	452
Zn (mg)	3	5
Vitamin C (mg)	61	122
Riboflavin (mg)	-	1
Niacin (mg)	1	8
Vitamin A (IU)	3128	23340

The aim of this study is to investigate the effects of rosemary and parsley at two levels (2 and 4g kg<sup>-1</sup> B.W.) on aflatoxicosis B<sub>1</sub> by fish, *Oreochromis niloticus* at 0.25 and 0.5 of the LD<sub>50</sub> of AFB<sub>1</sub>.

**MATERIALS AND METHODS**

**Preparation of Aflatoxin B<sub>1</sub>**

Aflatoxin B<sub>1</sub> was produced on liquid medium (Potato dextrose) by *Aspergillus parasiticus* (NRRL 2999) according to Ready *et al.* (1971). Aflatoxin B<sub>1</sub> was dissolved in chloroform and quantitatively estimated by thin layer chromatography, TLC (AOAC, 2000). So, chloroform was evaporated to dryness on a rotary vacuum evaporator at 40°C and redissolved in Dimethylsulfoxide (DMSO) 25% (1:3 water) to the requirement of each aflatoxin concentration. AFB<sub>1</sub> was freshly dissolved in DMSO before injection.

**Herbal Materials and Preparation of Their Extracts**

Fresh rosemary and parsley leaves were obtained from a local farm and carefully washed with tap water then left to dry in the dark at room temperature. Twenty gram of the ground leaves were extracted for 24 h by soaked in 500 ml of methanol (70%). The extract was then filtered and the filtrate was divided into two amounts (one part and its double) before evaporating them till dryness in a rotary evaporator (45°C). The residues of the two amounts were dissolved in constant volume of 25% DMSO to obtain the two concentrations of herbs extract. The dose levels of 0, 2 and 4 g kg<sup>-1</sup> B.W. were divided into double dose, the first was injected at the start of the experimental period and the second dose was injected one week later.

### Fish and Experimental Design

Two hundred and eighty eight fingerlings of *O. niloticus* were obtained from El-Serw fish farm, where this study was carried out in summer season 2007. The fish were acclimated to aquaria conditions for two weeks before the experiment was initiated. Six fish (approximately the same size, 20g average) were stocked into each of the 48 aquaria which contained 50 l of water, three glass aquaria (70X40X30 cm) for each treatment, the aquaria were provided with continuous aeration and their water was changed partially daily and totally weekly. All fish were received diet twice daily at a daily feeding rate of 3% of the actual body weight, six days weekly for two weeks. Fish were divided into 16 groups and were administered the test compounds interperitoneally (I.P.). Their effects were studied at the end of the 2<sup>nd</sup> week. The experimental setup used is shown in Table (1). AFB<sub>1</sub> was tested at three levels (0, 9 and 18 mg kg<sup>-1</sup> B.W.) being 0, 0.25 and 0.50 the LD<sub>50</sub>, respectively according to El-Barbary (2008) in a single dose, while both of rosemary and parsley extracts were used at three levels (being 0, 2 and 4 g kg<sup>-1</sup> B.W.) divided into 2 doses (pretreatment at the start of the experiment and one week later). AFB<sub>1</sub> and herbs' extracts were mixed together directly before administration.

**Table (1): Explanation of the experimental groups**

Groups	Pretreatment first week	Second week
A	Saline	Saline
B	DMSO 25%	DMSO 25%
F <sub>1</sub>	DMSO 25%	AFB <sub>1</sub> 9 mg/kg B.W.
F <sub>2</sub>	DMSO 25%	AFB <sub>1</sub> 18mg/kg B.W.
R <sub>1</sub>	Rosemary 1g/kg B.W.	Rosemary 1g/kg B.W.
F <sub>1</sub> R <sub>1</sub>	Rosemary 1g/kg B.W.	Rosemary 1g/kg B.W.+ AFB <sub>1</sub> 9 mg/kg B.W.
F <sub>2</sub> R <sub>1</sub>	Rosemary 1g/kg B.W.	Rosemary 1g/kg B.W.+ AFB <sub>1</sub> 18mg/kg B.W.
R <sub>2</sub>	Rosemary 2g/kg B.W.	Rosemary 2g/kg B.W.
F <sub>1</sub> R <sub>2</sub>	Rosemary 2g/kg B.W.	Rosemary 2g/kg B.W.+ AFB <sub>1</sub> 9 mg/kg B.W.
F <sub>2</sub> R <sub>2</sub>	Rosemary 2g/kg B.W.	Rosemary 2g/kg B.W.+ AFB <sub>1</sub> 18mg/kg B.W.
P <sub>1</sub>	Parsley 1g/kg B.W.	Parsley 1g/kg B.W.
F <sub>1</sub> P <sub>1</sub>	Parsley 1g/kg B.W.	Parsley 1g/kg B.W. + AFB <sub>1</sub> 9 mg/kg B.W.
F <sub>2</sub> P <sub>1</sub>	Parsley 1g/kg B.W.	Parsley 1g/kg B.W. + AFB <sub>1</sub> 18mg/kg B.W.
P <sub>2</sub>	Parsley 2g/kg B.W.	Parsley 2g/kg B.W.
F <sub>1</sub> P <sub>2</sub>	Parsley 2g/kg B.W.	Parsley 2g/kg B.W. + AFB <sub>1</sub> 9 mg/kg B.W.
F <sub>2</sub> P <sub>2</sub>	Parsley 2g/kg B.W.	Parsley 2g/kg B.W. + AFB <sub>1</sub> 18mg/kg B.W.

### Analytical Methods

At the end of the 2<sup>nd</sup> week of the experiment, blood samples were withdrawn from the fish heart of each group to determinate some blood parameters using commercial colorimetric kits (Diamond, Diagnostic, Egypt), and the obtained data were statistically analyzed by one way analysis of variance using a software (SAS, 1996). Three fish from each group were homogenized and prepared to determinate the residues of AFB<sub>1</sub> in fish by TLC (AOAC, 2000). The histological examination of the fish livers were performed after the preparation of livers which were dissected out from each group and fixed in 10% neutralized formalin solution until use according to the technique of Roberts (2001).

## RESULTS AND DISCUSSION

### Mortality Rate

The mortality rate caused by interperitoneal injection with AFB<sub>1</sub> alone or in combination with rosemary or parsley extracts was the highest in AFB<sub>1</sub> injected fish as shown in Table (2). The MR% gradually increased by increasing the AFB<sub>1</sub> level in all AFB<sub>1</sub> groups, while this increase in MR% was significantly reduced by using the herbs' extracts against AFB<sub>1</sub>. The positive effects of the herbs' extracts on MR% were observed. The level of both extracts 2g kg<sup>-1</sup> B.W. reflected the most significant decrease in MR% with the two levels of AFB<sub>1</sub> comparing with the level 4g kg<sup>-1</sup> B.W. The reduction in MR% ranged from 37.39, 33.30, 37.39 to 42.90% in groups F<sub>1</sub>R<sub>1</sub>, F<sub>2</sub>R<sub>1</sub>, F<sub>1</sub>P<sub>1</sub> and F<sub>2</sub>P<sub>1</sub>, respectively as compared with the AFB<sub>1</sub> groups (F<sub>1</sub> and F<sub>2</sub>). While, this reduction in MR% ranged

from 12.60 to 16.60% in groups F<sub>1</sub>R<sub>2</sub> and F<sub>2</sub>R<sub>1</sub>, respectively and 0 to 33.30% in F<sub>1</sub>P<sub>2</sub> and F<sub>2</sub>P<sub>2</sub> as compared to F<sub>1</sub> and F<sub>2</sub> groups, respectively.

**Table (2): Mortality rate (MR%) of Nile tilapia I.P. injected with AFB<sub>1</sub> with and without herbal plants extract (means ± standard errors).**

Groups	MR%*
A	5.5 <sup>e</sup> ± 5.5
B	5.5 <sup>e</sup> ± 5.5
F <sub>1</sub>	44.4 <sup>bc</sup> ± 5.5
F <sub>2</sub>	66.6 <sup>a</sup> ± 0.0
R <sub>1</sub>	5.5 <sup>e</sup> ± 5.5
F <sub>1</sub> R <sub>1</sub>	27.8 <sup>cd</sup> ± 5.5
F <sub>2</sub> R <sub>1</sub>	44.4 <sup>bc</sup> ± 5.5
R <sub>2</sub>	11.1 <sup>ed</sup> ± 5.5
F <sub>1</sub> R <sub>2</sub>	38.8 <sup>bc</sup> ± 5.5
F <sub>2</sub> R <sub>2</sub>	55.5 <sup>ab</sup> ± 11.0
P <sub>1</sub>	11.1 <sup>ed</sup> ± 5.5
F <sub>1</sub> P <sub>1</sub>	27.7 <sup>cd</sup> ± 5.5
F <sub>2</sub> P <sub>1</sub>	38.0 <sup>bc</sup> ± 5.5
P <sub>2</sub>	5.0 <sup>e</sup> ± 5.5
F <sub>1</sub> P <sub>2</sub>	44.0 <sup>bc</sup> ± 5.5
F <sub>2</sub> P <sub>2</sub>	44.0 <sup>bc</sup> ± 5.5

a - e: Means in the same column superscripted with different letters are significantly different at (P ≤ 0.001).

\* MR% = the beginning number of fish – the end number of the live fish X 100/ the beginning number of fish

These results agree with the results of AFB<sub>1</sub>-residues which confirmed that the level of both herbs' extracts 2g kg<sup>-1</sup> B.W. was more effective in reducing the AFB<sub>1</sub>-residues than the level 4g kg<sup>-1</sup> B.W. especially with the AFB<sub>1</sub> level 9 mg kg<sup>-1</sup> B.W. The significant effect of AFB<sub>1</sub> on MR% was confirmed in previous studies with Nile tilapia (Marzouk *et al.*, 1994; Hussein *et al.*, 2000; Abdelhamid *et al.*, 2002 a; Tuan *et al.*, 2002 and El-Barbary and El-Shaieb, 2006). The positive effect of parsley could be attributed to its medicinal property as anti-inflammation (Peter, 2001). Also, both of rosemary and parsley have antioxidant properties due to polyphenolic compounds including ferulic acid and syringic acid that are the major phenolic compounds in parsley (El-Barbary, 2008). Ferulic acid exhibit a wide range of pharmacological effects including antiageing, anti-inflammatory, anticancer, antiapoptotic, antidiabetic and neuroprotective (Srinivasan *et al.*, 2006).

#### Quantitative Estimation of AFB<sub>1</sub> Residues

The residual analysis of AFB<sub>1</sub> in the whole body of fish which were injected with 9 mg AFB<sub>1</sub> kg<sup>-1</sup> B.W. with or without herbs' extracts (F<sub>1</sub>, F<sub>1</sub>P<sub>2</sub>, F<sub>1</sub>R<sub>2</sub>, F<sub>1</sub>P<sub>1</sub> and F<sub>1</sub>R<sub>1</sub>) (Fig. 1) showed that all these groups revealed the presence of AFB<sub>1</sub> residues (1.6 to 7.5 ppb) which consider less than the permissible limit (20 ppb) that was recommended by WHO (Diener *et al.*, 1985). In fish groups injected with AFB<sub>1</sub> with herbs' extracts, traces of AFB<sub>1</sub> were detected in their body at concentration ranged from 1.6 to 3.1 ppb.

Fig. (2) illustrates AFB<sub>1</sub> residues of the fish groups injected with the level of AFB<sub>1</sub> 18 mg kg<sup>-1</sup> B.W. with or without herbs' extracts (F<sub>2</sub>, F<sub>2</sub>P<sub>1</sub>, F<sub>2</sub>R<sub>1</sub>, F<sub>2</sub>P<sub>2</sub> and F<sub>2</sub>R<sub>2</sub>) which, showed contrast results to those in Fig. (1), since there were no-residues of AFB<sub>1</sub> in either F<sub>2</sub> (18 mg AFB<sub>1</sub> kg<sup>-1</sup> B.W.) or F<sub>2</sub>P<sub>1</sub> (18 mg AFB<sub>1</sub> + 2g parsley kg<sup>-1</sup> B.W.). Whereas AFB<sub>1</sub>-residues ranged from 2.1, 9.6 to 23.0 ppb in F<sub>2</sub>R<sub>1</sub>, F<sub>2</sub>P<sub>2</sub> and F<sub>2</sub>R<sub>2</sub>, respectively.

These results indicate that the herbs' extracts have potency for reducing the AFB<sub>1</sub> residues, particularly in cases of the low levels of AFB<sub>1</sub> and the herbs' extracts. Also, parsley could have higher potency than rosemary.

The absence of AFB<sub>1</sub> residues in F<sub>2</sub> group (18 mg AFB<sub>1</sub> kg<sup>-1</sup> B.W.) could be attributed to the high level of AFB<sub>1</sub> which acts as an acute dose, so its metabolism may be rapid and severely converted into other metabolites which could have more toxicity than AFB<sub>1</sub>.

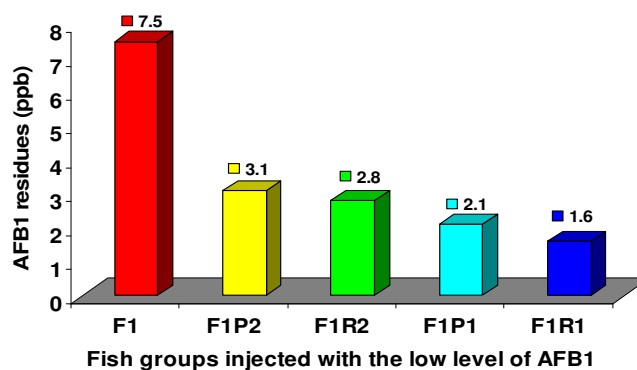


Fig. (1): AFB<sub>1</sub> residues in the experimented fish injected with the low dose of AFB<sub>1</sub>.

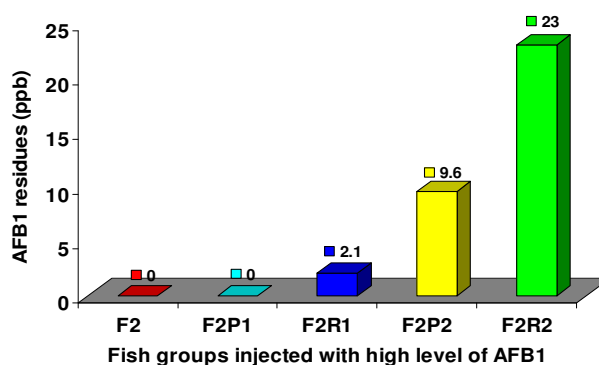


Fig. (2): AFB<sub>1</sub> residues in the experimented fish injected with the high dose of AFB<sub>1</sub>.

The negative results of both MR% and histopathological examination in F<sub>2</sub> group may confirm that. On the other hand, the low level of AFB<sub>1</sub> acts as subacute dose; so, its metabolism may be gradual and slight so its MR% and histopathological manifestation had better results comparing with F<sub>2</sub> group. The mechanism of parsley and rosemary extract may be due to inhibition of the metabolic activation through inhibition of cytochrome P<sub>450</sub> enzymes, and also through induction of the detoxifying enzyme glutathione S-transferase. In this respect, similar results were recorded by Soliman *et al.* (1998 and 2000). Yet, Abdelhamid *et al.* (2004 b and 2007) recorded high level of AFB<sub>1</sub> in whole body of Nile tilapia fish. On the other hand, Abdelhamid *et al.* (2002 a, 2002 b and 2004 a) reported that there were no AFB<sub>1</sub> residues in *O. niloticus* body. These variable results may be due to AFB<sub>1</sub> level and exposure time as well as to sensitivity variation among fish species to AFB<sub>1</sub>. Recently, Oliveira and Furlong (2008) reported that phenolic extracts of different edible plants have antifungal and antimycotoxigenic activity.

#### Blood Parameters

There were significant reduction in Hb, TP and Gl levels and increase in the albumin (Al) concentration and activity of hepatic transaminase enzymes (AST and ALT) at all AFB<sub>1</sub> groups, whether injected with AFB<sub>1</sub> alone or AFB<sub>1</sub> with herbs' extracts comparing to the control group (A). Table (3) indicates that the control solvent (B) showed approximately similar values of most blood parameters when compared to the control (A). That indicates the absence of toxicity due to DMSO administration. This alteration in haematology and biochemistry of blood was gradually increased by increasing the level of AFB<sub>1</sub>. On contrary, these negative alterations due to AFB<sub>1</sub> were significantly improved by using rosemary and parsley extracts, particularly at the low level of AFB<sub>1</sub>.

However, these improved values remained lower than the control values. Also, the study indicates that R<sub>1</sub> and P<sub>1</sub> showed significant alterations in some of the studied blood parameters when compared to the control.

**Table (3): The influence of AFB<sub>1</sub> with or without either of rosemary or parsley extract on some blood parameters of *O. niloticus* ( $\bar{X} \pm SE$ )**

Groups	Hb (g dl <sup>-1</sup> )	TP (g dl <sup>-1</sup> )	AL (g dl <sup>-1</sup> )	GL (g dl <sup>-1</sup> )	AST U/I	ALT U/I
A	6.74 <sup>a</sup> ±0.04	4.31 <sup>b</sup> ±0.06	1.35 <sup>c</sup> ±0.01	2.96 <sup>a</sup> ±0.05	56.37 <sup>ef</sup> ±0.33	22.47 <sup>ef</sup> ±0.23
B	6.48 <sup>ab</sup> ±0.06	4.38 <sup>a</sup> ±0.05	1.40 <sup>d</sup> ±0.01	2.98 <sup>a</sup> ±0.06	56.40 <sup>ef</sup> ±0.35	21.53 <sup>gh</sup> ±0.40
F <sub>1</sub>	5.36 <sup>f</sup> ±0.03	3.71 <sup>g</sup> ±0.05	1.67 <sup>b</sup> ±0.02	2.05 <sup>g</sup> ±0.07	69.37 <sup>a</sup> ±1.30	24.01 <sup>d</sup> ±0.13
F <sub>2</sub>	4.60 <sup>g</sup> ±0.20	3.10 <sup>i</sup> ±0.01	1.51 <sup>c</sup> ±0.04	1.59 <sup>h</sup> ±0.11	70.23 <sup>a</sup> ±0.55	28.80 <sup>a</sup> ±0.06
R <sub>1</sub>	6.32 <sup>bc</sup> ±0.11	4.04 <sup>d</sup> ±0.08	1.37 <sup>e</sup> ±0.02	2.68 <sup>c</sup> ±0.11	55.20 <sup>f</sup> ±0.60	22.07 <sup>fg</sup> ±0.09
F <sub>1</sub> R <sub>1</sub>	5.25 <sup>f</sup> ±0.04	3.75 <sup>g</sup> ±0.02	1.53 <sup>c</sup> ±0.07	2.22 <sup>f</sup> ±0.08	63.40 <sup>cd</sup> ±0.90	24.03 <sup>d</sup> ±0.12
F <sub>2</sub> R <sub>1</sub>	4.89 <sup>g</sup> ±0.01	3.97 <sup>d</sup> ±0.03	1.42 <sup>d</sup> ±0.01	2.55 <sup>d</sup> ±0.01	61.47 <sup>d</sup> ±0.86	29.13 <sup>a</sup> ±0.03
R <sub>2</sub>	6.12 <sup>cd</sup> ±0.39	4.42 <sup>a</sup> ±0.02	1.64 <sup>b</sup> ±0.03	2.77 <sup>b</sup> ±0.02	56.13 <sup>ef</sup> ±1.01	20.67 <sup>i</sup> ±0.12
F <sub>1</sub> R <sub>2</sub>	5.95 <sup>d</sup> ±0.05	4.22 <sup>c</sup> ±0.01	1.49 <sup>c</sup> ±0.03	2.73 <sup>b</sup> ±0.03	58.47 <sup>e</sup> ±0.37	24.00 <sup>d</sup> ±0.31
F <sub>2</sub> R <sub>2</sub>	4.77 <sup>g</sup> ±0.10	3.56 <sup>h</sup> ±0.03	1.45 <sup>cd</sup> ±0.03	2.10 <sup>g</sup> ±0.03	58.13 <sup>e</sup> ±0.70	23.00 <sup>e</sup> ±0.17
P <sub>1</sub>	6.32 <sup>bc</sup> ±0.49	4.05 <sup>d</sup> ±0.02	1.36 <sup>de</sup> ±0.01	2.69 <sup>c</sup> ±0.03	58.33 <sup>e</sup> ±1.22	21.37 <sup>h</sup> ±0.01
F <sub>1</sub> P <sub>1</sub>	5.90 <sup>e</sup> ±0.05	3.72 <sup>g</sup> ±0.07	1.51 <sup>c</sup> ±0.03	2.21 <sup>ef</sup> ±0.07	66.27 <sup>b</sup> ±0.50	25.84 <sup>c</sup> ±0.04
F <sub>2</sub> P <sub>1</sub>	4.68 <sup>g</sup> ±0.06	3.82 <sup>fg</sup> ±0.10	1.53 <sup>c</sup> ±0.01	2.28 <sup>e</sup> ±0.02	61.20 <sup>d</sup> ±0.73	26.55 <sup>b</sup> ±0.33
P <sub>2</sub>	6.28 <sup>bcd</sup> ±0.04	4.04 <sup>d</sup> ±0.03	1.33 <sup>c</sup> ±0.03	2.71 <sup>b</sup> ±0.06	57.90 <sup>e</sup> ±0.87	22.05 <sup>fg</sup> ±0.05
F <sub>1</sub> P <sub>2</sub>	5.41 <sup>f</sup> ±0.05	3.89 <sup>e</sup> ±0.02	1.61 <sup>b</sup> ±0.01	2.28 <sup>e</sup> ±0.03	66.77 <sup>b</sup> ±0.99	23.01 <sup>e</sup> ±0.15
F <sub>2</sub> P <sub>2</sub>	4.56 <sup>g</sup> ±0.14	3.90 <sup>de</sup> ±0.02	1.92 <sup>a</sup> ±0.02	1.98 <sup>g</sup> ±0.04	64.77 <sup>bc</sup> ±1.10	24.06 <sup>d</sup> ±0.09

a - i: Means in the same column superscripted with different letters are significantly different at ( $P \leq 0.001$ ).

So, the positive effects of the two levels of these extracts were clearly observed with the low level than the high level of AFB<sub>1</sub>. While, the high level of rosemary and parsley was better with the high level of AFB<sub>1</sub>. These alterations in blood parameters among fish groups may be due to the alterations in histological structure of livers of AFB<sub>1</sub>-injected fish leading to inhibition of blood synthesis, where liver plays an important role in this process. Similarly, Abdelhamid *et al.* (2002 a) and El-Barbary and El-Shaieb (2006) reported that AFB<sub>1</sub> reduced TP, Al and Gl of *O. niloticus*. In the same trend, Abdelhamid *et al.* (2007) found that AFB<sub>1</sub> caused significant decrease in TP, Al and Gl of aflatoxicated *O. niloticus* fish.

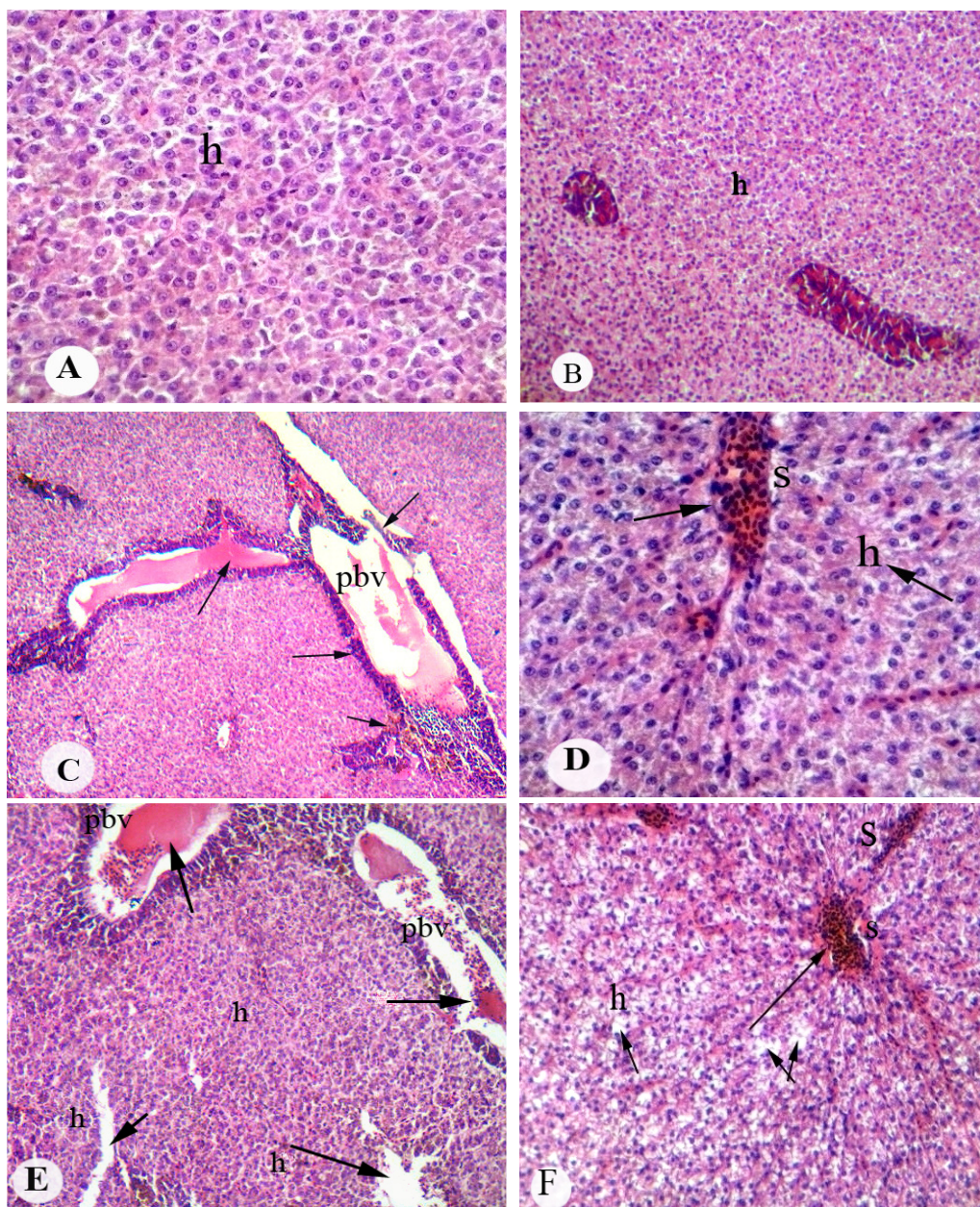
This reduction in TP levels may be due to the hepatotoxic effect of AFB<sub>1</sub>. Whereas the reduced Gl levels in AFB<sub>1</sub> injected fish may have been the result of lymphocytolysis (Sahoo *et al.*, 1998). While Youssef and Ashry (1999) attributed the increase in activity of AST and ALT enzymes to the hepatotoxic effect of AFB<sub>1</sub> and consequently hepatic cell damage and liver dysfunction.

#### Clinical and Histopathological Findings

The present study revealed that the toxicity signs began to exist together with mortalities. The most common clinical signs observed were lethargy, loss of appetite, sluggish movement, dark discoloration of the skin and respiratory manifestations. Macroscopically, the common lesions in all necropsied aflatoxicated fish were accumulation of fluids in abdomen, congested gills and dark liver. These symptoms were varied according to the treatment. The photomicrograph of liver showing microscopically the parenchymal architecture of hepatocytes in the control group (A) with central nuclei (Fig. 3A). Fish of the solvent control group (B) presented normal structure of the liver (Fig. 3B). Also, no clear histological changes were observed in any of fish groups which injected with herbs' extracts alone at the different levels (R<sub>1</sub>, R<sub>2</sub>, P<sub>1</sub>, P<sub>2</sub>). In contrast, fish injected with AFB<sub>1</sub> presented pathological changes in hepatopancreas which increased in severity with increasing the level (groups F<sub>1</sub>, F<sub>2</sub> at 9 and 18 mg kg<sup>-1</sup> B.W respectively). These changes in F<sub>1</sub> group were severe hemolysis in the portal blood vessels (PBV) and presence melanomacrophages (MMC) (Fig. 3C). In addition, lysis of hepatocyte membranes (necrotic cells) besides dilation and congestion in blood sinusoid were noticed (Fig. 3D); while the lesions among F<sub>2</sub> were severe hemolysis, dilation in the portal blood vessels and large area of degenerated hepatocytes (Fig. 3E). Also, liver shows diffuse vaculation and necrosis of hepatocytes and nuclei displaced to the cell periphery; in addition to, congestion and dilation in blood sinusoid (Fig. 3F), thrombosis formation in blood vessels (Fig. 4G) and accumulation of MMC and hemosiderin besides coagulative necrosis in hepatocytes were recorded too (Fig. 4H). These severe pathological alterations in hepatopancrease caused by AFB<sub>1</sub> alone became less severe when these fish were injected with AFB<sub>1</sub> plus either rosemary or parsley at the different levels. The effects of the low level of rosemary on the two levels of AFB<sub>1</sub> were observed in Figs. 4 I and J; whereas, group F<sub>1</sub>R<sub>1</sub> showed slight



degeneration of hepatocytes (Fig. 4I), while the lesions in group F<sub>2</sub>R<sub>1</sub> were necrosis in hepatocytes besides congestion of some pancreatic acini (Fig. 4J).

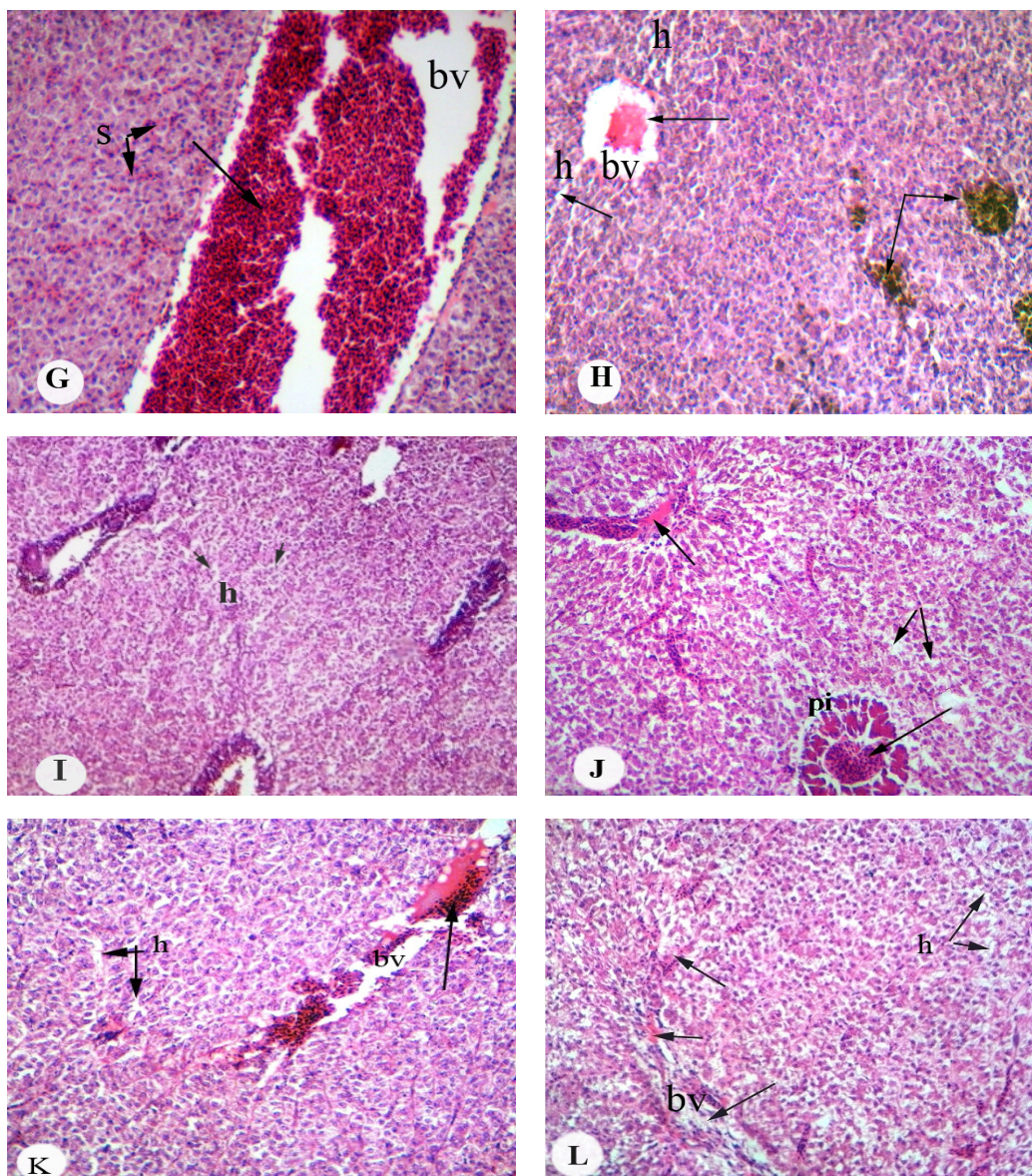


**Fig. 3:** Histopathological changes in liver of *O niloticus* injected with different levels of AFB<sub>1</sub> as compared to control (stained with H&E). (A&B): The control and control solvent fish groups showing normal structure of the liver (X600&250, respectively). (C&D); fish injected with AFB<sub>1</sub> (9 mg kg<sup>-1</sup> B.W., F<sub>1</sub> group) showing hemolysis and dilation in pbv (C, X200) besides congestion and dilation in sinusoid with necrosis (D, X600). (E&F); fish injected with AFB<sub>1</sub> (18 mg kg<sup>-1</sup> B.W., F<sub>2</sub> group) showing dilation and hemolysis in pbv, degeneration in hepatocytes (E, X300), vacuolation and necrosis in hepatocytes and congestion with dilation in sinusoid (F, X450). h; hepatocyte. s; sinusoid. pbv; portal blood vessel.

The effects of the high level of rosemary with AFB<sub>1</sub> were observed, where the hepatocytes showed condensed cytoplasm and loss of contact between hepatocytes (Figs. 4 K & L), in addition to the presence of few numbers of inflammatory cells (Fig. 4L), besides mild congestion and hemolysis in blood vessels and necrosis of hepatocytes (Fig. 4K). Concerning the effect of the low level of parsley with different levels of AFB<sub>1</sub>, it may have a positive effect in ameliorating the toxicity of AFB<sub>1</sub>, since the liver of group (F<sub>1</sub>P<sub>1</sub>) showed normal structure of hepatocytes with slight hemolysis of hepatic acini and presence of few numbers of MMC (Fig. 5M). Similar



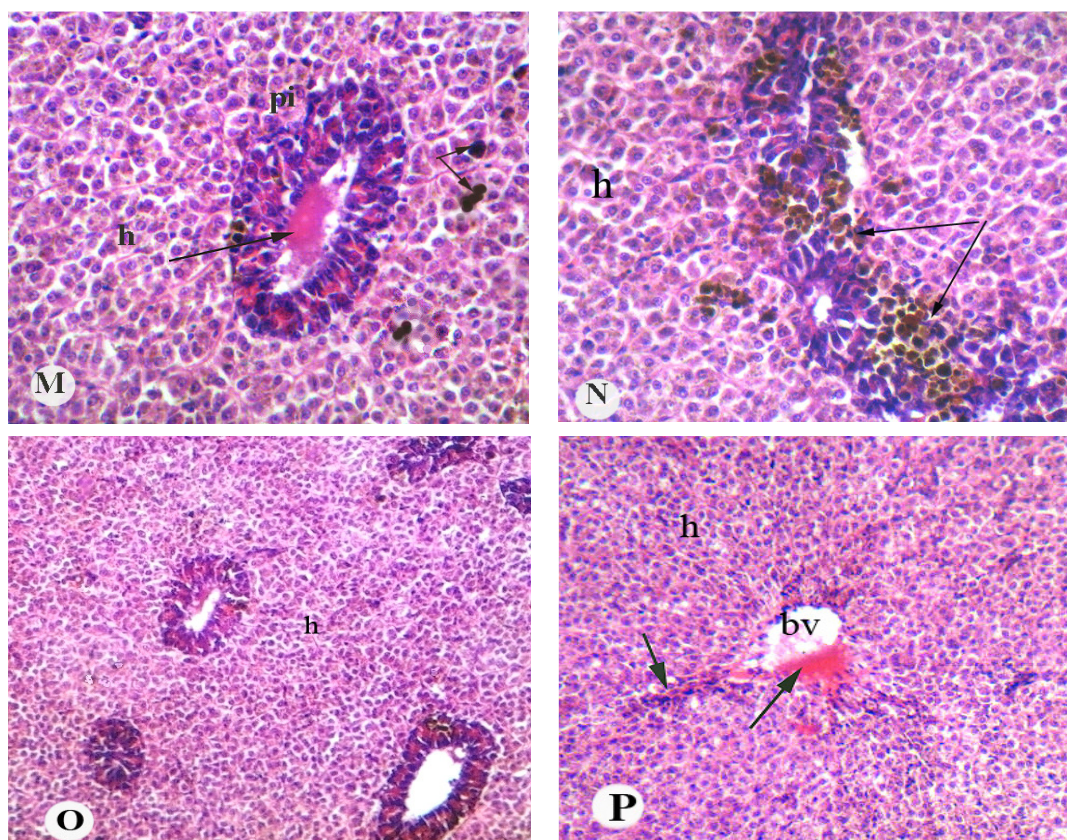
findings were observed in group F<sub>2</sub>P<sub>1</sub> but with diffuse of hemosiderin and MMC (Fig. 5N). The effect of the high level of parsley against AFB<sub>1</sub> was presented in groups F<sub>1</sub>P<sub>2</sub> and F<sub>2</sub>P<sub>2</sub>, since the liver of group F<sub>1</sub>P<sub>2</sub> showed normal hepatocytes (Fig. 5O). While group F<sub>2</sub>P<sub>2</sub> showed slight hemolysis and presence of few inflammatory cells (Fig. 5P). Similar hepatic lesions were reported by El-Banna *et al.* (1992), Hussein *et al.* (2000) and Abdelhamid *et al.* (2002 a) who described congestion, vacuolar degeneration of hepatocytes and activation of melanomacrophages in aflatoxicated tilapia. Also, El-Barbary and El-Shaieb (2006) reported that the liver of aflatoxicated fish showed severe vacuolation of hepatocytes of mainly fatty changes besides focal coagulative necrosis, focal replacements of the hepatic parenchyma with extravagated blood, and severe congestion and hemorrhage. In the same trend, also Mehrim *et al.*, (2006) emphasized similar clinical sings and histopathological lesions in the liver of aflatoxicated Nile tilapia.



**Fig. 4:** Histopathological changes in liver of *O niloticus* injected with AFB<sub>1</sub> with or without rosemary at the different levels (stained with H&E). (G, H); fish injected with (18 mg kg<sup>-1</sup> B.W., F<sub>2</sub> group) showing thrombosis formation in bv (G, X250), coagulative necrosis, besides diffusion of MMC and hemosiderin (H, X250). (I); fish injected with AFB<sub>1</sub> + rosemary (9 mg and 2g kg<sup>-1</sup> B.W., F<sub>1</sub>R<sub>1</sub> group) showing slight necrosis (X250). (J); fish injected with AFB<sub>1</sub> + rosemary (18 mg and 2g kg<sup>-1</sup> B.W., F<sub>2</sub>R<sub>1</sub> group) showing congestion and necrosis in pi (X400). (K); fish injected with AFB<sub>1</sub> + rosemary (9 mg and 4g kg<sup>-1</sup> B.W., F<sub>1</sub>R<sub>2</sub> group) showing necrosis and congestion (X400). (L); fish injected with AFB<sub>1</sub> + rosemary (18 mg and 4g kg<sup>-1</sup> B.W., F<sub>2</sub>R<sub>2</sub> group) showing necrosis and congestion (X400). bv; blood vessel. pi; pancreatic acini.



The positive effects of both of the parsley and rosemary on overcoming the toxic effects of AFB<sub>1</sub> could be attributed to the antioxidative and nutritive properties of these herbs. These results showed that the ability of parsley extract to counteract the toxic effects of AFB<sub>1</sub> on the fish could be better than rosemary extract. That may be attributed to the high nutritive value of parsley that included too high percent of vitamins (A, C, riboflavin and niacin) and minerals (Fe, Mg, P, K, Ca, Na and Zn) compared to rosemary. Vitamin C is related to the immunological system performance, and has antioxidant properties. This antioxidant activity of Vit. C makes it as a hunter of free radicals, thus preventing the autointoxication of immunological cells, such as macrophages which are the first processors of the information about the alien bodies and maximizing the defense of fish (Brake, 1997). Also, metal ions such as Se, Zn, Cu, Mn and Fe are essential for most organisms. Essential trace elements are important parts of antioxidant enzymes as superoxide dismutase and glutathione peroxidase and may affect the antioxidant defense system (Hung *et al.*, 2007).



**Fig. 5:** Histopathological changes in liver of *O niloticus* injected with AFB<sub>1</sub> + parsley at the different levels (stained with H&E). (M); fish injected with AFB<sub>1</sub>+ parsley (9 mg and 2g kg<sup>-1</sup> B.W., F<sub>1</sub>P<sub>1</sub> group) showing hemolysis in pi and presence of MMC (X600). (N); fish injected with AFB<sub>1</sub>+ parsley (18 mg and 2g kg<sup>-1</sup> B.W., F<sub>2</sub>P<sub>1</sub> group) showing diffusion of MMC and hemosiderin (X600).(O); fish injected with AFB<sub>1</sub>+ parsley (9 mg and 4g kg<sup>-1</sup> B.W., F<sub>1</sub>P<sub>2</sub> group) showing normal structure (X250). (P); fish injected with AFB<sub>1</sub>+ parsley (18 mg and 4g kg<sup>-1</sup> B.W., F<sub>2</sub>P<sub>2</sub> group) showing slight hemolysis and infiltration (X400).

## CONCLUSIONS

From previous results, it could be recommended the useful using of both of medicinal herbs, namely rosemary and parsley, at low level (2 g kg<sup>-1</sup> B.W.) to eliminate the drastic effects of aflatoxin B<sub>1</sub> on *O. niloticus*. Yet, it must be exist more scientific efforts to use the medicinal herbs and other natural materials against the contamination with the mycotoxins. But, it will be still the prevention from toxic effects of aflatoxin B<sub>1</sub> and other toxins are more useful usually.

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#### **REFERNCES**

- Abdelhamid, A.M., A.I. Mehrim, and F.F. Khalil, 2004 a. Detoxification of aflatoxin–contaminated diet of tilapia fish using dietary supplementation with egg shell, Betafin, clay or silica. J. Agric. Sci. Mansoura Univ., 29: 3163-3174. (ISSN: 1110-0346).
- Abdelhamid, A.M., A.E. Abdel- Khalek, A.I. Mehrim, and F.F. Khalil, 2004 b. An attempt to alleviate aflatoxicosis on Nile tilapia fish by dietary supplementations with chicken-hatchery by-products (egg shells) and shrimp processing wastes (shrimp shells). 2- On clinical, blood and histological parameters. J. Agric. Sci. Mansoura Univ., 29 : 6175 - 6196. (ISSN: 1110-0346)
- Abdelhamid, A.M., F.F. Khalil, M.I. El-Barbary, V.H. Zaki, and H.S. Hussein, 2002 a. Feeding Nile tilapia on Biogen® to detoxify aflatoxic diets. Proc.1<sup>st</sup> Conf. Animal & Fish Prod., Mansoura, 24&25, Sept., pp: 207-230. (ISSN: 1110-0346)
- Abdelhamid, A.M., F.I. Magouz, M.F.E. Salem, A.A. Mohamed and M.K. Mohsen, 2002 b. Effect of graded levels of aflatoxin B<sub>1</sub> on growth performance and biochemical, chromosomal and histological behaviour of Nile tilapia *Oreochromis niloticus*. Proc.1<sup>st</sup> Conf. Animal & Fish Prod., Mansoura, 24&25, Sept., pp:231-250. (ISSN: 1110-0346)
- Abdelhamid, A.M., M.F.I. Salem, A.I. Mehrim, and El-Sharawy, M.A.M. 2007. Nutritious attempts to detoxify aflatoxic diets of tilapia fish: 1-Fish performance, feed and nutrients utilization, organs indices, residues and blood parameters. Egyptian J. Nutrition and Feeds, 10: 205-223. (ISSN: 1110-6360).
- AOAC, 2000. Association of Official Analytical Chemists. Official Methods of Analysis, 17<sup>th</sup> Ed. Washington, DC.
- Brake, L., 1997. Immune status of vitamins. Feed Mix., 5: 21-24.
- Caillet, S., H. Yu, S. Lessard, G. Lamoureux, D. Ajdukovic, and M. Lacroix, 2007. Fenton reaction applied for screening natural antioxidants. Food Chemistry, 100: 542-552. (DOI:10.1016/j.foodchem.2005.10.009).
- Diener, U.L., N.D. Davis, and D.A. Danilson, 1985. Detoxification of aflatoxin contaminated corn makes grain safe for feeding. Alabama Agricultural Experiment Station. Highlights of Agricultural Research (Auburn University). 32: 15.
- El-Banna, R., H.M. Teleb, M.M. Hadi, and F.M. Fakhry, 1992. Performance and tissue residue of tilapias fed dietary aflatoxin. J. Veterinary Medical Giza, 40: 17-23. (ISSN: 1110-7219).
- El-Barbary, M.I., 2008. Aflatoxin B<sub>1</sub> induced-changes in protein electrophoretic pattern and DNA in *Oreochromis niloticus* with special emphasis on the protective effect of rosemary and parsley extracts. (Under publication in J. of Agri. Envir. Sci.).

- El-Barbary, M.I. and A.F. El-Shaieb, 2006. A contribution on the role of vitamin C in *Oreochromis niloticus* fed on diets containing aflatoxin B<sub>1</sub> and/or *Aspergillus parasiticus* fungus. Egyptian Journal of Aquatic research, 32: 425-442. (ISSN: 1110-0354).
- Farrel, K.T., 1990. Spices, Candiments and Seasonings. 2<sup>nd</sup> edition. AV 1 book, Van Nostrand Reinhold, New York. (ISBN-13: 978-0412122811).
- Gyamfi, M.A. and Y. Aniya, 1998. Medicinal herb, *Thonningia sanguinea* protects against aflatoxin B<sub>1</sub> acute hepatotoxicity in Fischer 344 rats. Hum. Exp. Toxicol., 17: 418-423. (DOI: 10.1177/096032719801700802).
- Gyamfi, M.A., M. Yonamine, and Y. Aniya, 1999. Free radical scavenging action of medicinal herbs from Ghana *Thonningia sanguinea* on experimentally induced liver injuries. Gem. Pharm., 32: 661-667. (PMID: 10401991 [PubMed - indexed for MEDLINE])
- Hinneburg, I., H.J.D. Dorman, and R. Hiltunen, 2006. Antioxidant activities of extracts from selected culinary herbs and spices. Food Chemistry, 97: 122-129. (<http://cat.inist.fr/?aModele=afficheN&cpsid=17508689>), (ISSN: 0308-8146).
- Huang, M.T., C.T. Ho, Z.Y. Wang, T. Ferraro, Y.R. Lou, K. Stauber, W. Ma, C. Gerogiadis, J.D. Laskin, and A.H. Conney, 1994. Inhibition of skin tumorigenesis by rosemary and its constituents carnosol and ursolic acid. Cancer Res., 54: 701-708. (<http://cancerres.aacrjournals.org/cgi/content/abstract/54/3/701>).
- Hung, S.W., C. Yu, and W. Wang, 2007. *In vivo* effects of adding singular or combined anti-oxidative vitamins and/or minerals to diets on the immune system of tilapia (*Oreochromis hybrids*) Peripheral blood monocyte-derived, anterior kidney-derived, and spleen-derived macrophages. Vet. Immunol., Immunopathol., 115: 87-99. (<http://cat.inist.fr/?aModele=afficheN&cpsid=18427551>), (ISSN: 0165-2427).
- Hussein, S.Y., I.A.A. Mekkawy, Z.Z. Moktar, and M. Mubarak, 2000. Protective effect of *Nigella sativa* seed against aflatoxicosis in *Oreochromis niloticus*. Proc. Conf. Mycotoxins and Dioxins and the Environment, Bydgoszcz, 25 – 27 Sept., pp: 109 – 130. (ISBN: 83-912646-0-2).
- IARC., 1993. IARC monographs on the evaluation of carcinogenic risks to humans. v.56: Some naturally occurring substances: Food items and constituents, heterocyclic aromatic amines and mycotoxins. Proceedings of IARC Working Group on the Evaluation of Carcinogenic Risks to Humans, Jun. 9-16, 1992, IARC, Lyon France, pp: 245-395.
- Lampe, J.W., I. B. King, S. Li, M.T. Grate, K.V. Barale, C. Chen, Z. Feng, and J.D. Potter, 2000. Brassica vegetables increase and apiaceous vegetables decrease cytochrome P<sub>450</sub> 1A2 activity in humans: Changes in caffeine metabolite ratios in response to controlled vegetable diets. Carcinogenesis, 21: 1157-1162. (<http://carcin.oxfordjournals.org/cgi/content/abstract/21/6/1157>).
- Marzouk, M.S., M.M. Bashandi, R. El-Danna, M. Moustafa, and M.A. Eissa, 1994. Hematological studies on aflatoxicosis. Egyptian Journal of Comparative Pathology and Clinical Pathology, 7: 497-504. (ISSN: 1110-7537).
- Mehrim, A. I., A. M. Abdelhamid, A. Abou-Shousha, M.F.I. Salem, and M.A.M.M. El-Sharawy, 2006. Nutritious attempts to detoxify aflatoxic diets of tilapia fish: 2- Clinical, biochemical and histological parameters. Journal of the Arabian Aquaculture Society, 1: 69-90. (Issued at No.:1633/2004).

- Offord, E.A., K. Mace, O. Avant, and A.M.A. Pfefer, 1997. Mechanisms involved in the chemoprotective effects of rosemary extract studied in human liver and branchial cells. *Cancer Letters*, 114: 275-281. (DOI: 10.1016/S0304-3835(97)04680-6).
- Oliveira, M.D.S. and E.B. Furlong, 2008. Screening of antifungal and antimycotoxigenic activity of plant phenolic extracts. *World Mycotoxin Journal*, 1: 139-146. (DOI: 10.3920/WMJ2008.1006).
- Peter, K.V., 2001. Hand book of herbs and spices. Woodhead publishing Limited, Abington. (ISBN: 185573 5628).
- Rastogi, S., Y. Shukla, B.N. Paul, D.K. Chowdhuri, S.K. Khanna, and M. Das, 2007. Protective effect of *Ocimum sanctum* on 3-methylcholanthrene, 7,12-dimethylbenz (a) anthracene and aflatoxin B<sub>1</sub> induced skin tumorigenesis in mice. *Toxicology and Applied Pharmacology*, 224: 228-240. (DOI: 10.1016/j.taap.2007.05.020).
- Ready, T.V., L. Viswanathan, and T.A. Venkitesubramanian, 1971. High aflatoxin production on chemically defined medium. *App. Microbiol.*, 22: 393-396. (<http://aem.asm.org/cgi/content/abstract/22/3/393>).
- Roberts, R. J., 2001. *Fish Pathology*, 3<sup>rd</sup> edition, W.B. Saunders. (ISBN: 0-7020-2563-1).
- Sacan-Ozsoy, O., R. Yanardag, H. Orak, Y. Ozgey, A. Yarat, and Tunali, T. 2006. Effects of parsley (*Petroselinum crispum*) extract versus glibornuride on the liver of streptozotocin- induced diabetic rats. *J. of Ethnopharmacology*, 104: 175-181. (DOI: 10.1016/j.jep.2005.08.069 ).
- Sahoo, P.K., S.C. Mukherjee, A.K. Jain, and A. Mukherjee, 1998. Light and ultrastructural changes in opisthonephros of rohu, *Labeo rohita* during acute and subchronic aflatoxin B<sub>1</sub> toxicity (abstract). In: International Conference on Fisheries and Food Security beyond the year 2000. Chiang Mai, Thailand, p. 212.
- SAS, 1996. SAS Users Guid: Statistics. Version 2,5 Edition. SAS Institute, Inc., Cary, NC. . ([www.informatik.uni-trier.de/~ley/db/conf/sas/sas98.html](http://www.informatik.uni-trier.de/~ley/db/conf/sas/sas98.html)15 - k),(ISBN: 3-540-65014-8).
- Shen, H.M., C.Y. Shi, Y. Shen, and C.N. Ong, 1996. Detection of elevated reactive oxygen species level in cultured rat hepatocytes treated with aflatoxin B<sub>1</sub>. *Free. Radic. Biol. Med.*, 21: 139-146. (<http://www.ncbi.nlm.nih.gov/pubmed/8818628>), (PMID: 8818628 [PubMed - indexed for MEDLINE]).
- Singletary, K.W. and J.M. Nelshoppen, 1991. Inhibition of 7,12 dimethylbenz (a) anthracene (DMBA)-induced mammary tumorigenesis and *in vivo* formation of mammary DMBA-DNA adducts by rosemary extract. *Cancer Let.*, 60: 169-175. (<http://www.ncbi.nlm.nih.gov/pubmed/1933840>), (PMID: 1933840 [PubMed - indexed for MEDLINE]).
- Soliman, K. M., A.M. Ayesh, M. A. M. Essa, and K. Naguib, 1998. Fix-A-tox in aquaculture: 1-Effect of aflatoxin decontamination by a selective chemisorbent materials on the *Oreochromis niloticus* with considering fish processing efficiency. *J. Egypt. Ger. Soc. Zoot.*, 25 (A) Comparative Physiology, 1 – 19. (ISSN: 1110-5321).
- Soliman, M.K., R.H. Khalil, S.A. Youssef, and N.M. Mahfouz, 2000. Aflatoxicosis among cultured freshwater fish. Abstracts AQUA-2000, Nice-France, May 2 – 6, p: 801.
- Srinivasan, M., A.R. Sudheer, K.R. Pillai, P.R. Kumar, P.R. Sudhakaran, and V.P. Menon, 2006. Influence of ferulic acid on Y-radiation induced DNA damage, lipid peroxidation and antioxidant status in primary culture of isolated rat hepatocytes. *Toxicology*, 228: 249-258. (DOI: 10.1016/j.tox.2006.09.004).



- Tokuda, H., H. Ohigashi, K. Koshimizu, and Y. Ito, 1986. Inhibitory effects of ursolic acid and oleanic acid on skin tumor promotion by 12-O- tetradecanoylphorbol-13- acetate. *Cancer Let.*, 33: 279-285. (<http://www.ncbi.nlm.nih.gov/pubmed/3802058?dopt=Abstract>), (PMID: 3802058 [PubMed - indexed for MEDLINE]).
- Tuan, N.A., J. M. Grizzle, R. T. Lovell, B. B. Manning, and E.G. Rottinghaus, 2002. Growth and hepatic lesions of Nile tilapia *Oreochromis niloticus* fed diets containing aflatoxin B<sub>1</sub>. *Aquaculture*, 212: 311-319. (DOI: 10.1016/S0044-8486(02)00021-2).
- Vinitketkumnuen, U., T. Chewonarin, P. Dhumtanom, N. Lertprasertsuk, and C.P. Wild, 1999. Aflatoxin-albumin adduct formation after single and multiple dose of aflatoxin B<sub>1</sub> in rats treated with thai medicinal plants. *Mutation Research*, 428: 345-351. (DOI: 10.1016/S1383-5742(99)00060-5).
- Wong, B.Y., B.H. Lau, T. Yamasaki, and R.W. Teel, 1993. Inhibition of dexamethasone - induced cytochrome P<sub>450</sub> - mediated mutagenicity and metabolism of aflatoxin B<sub>1</sub> by Chinese medicinal herbs. *Eur. J. Cancer Prev.*, 2: 351 - 356. (<http://www.ncbi.nlm.nih.gov/sites/entrez>), (PMID: 8358288 [PubMed - indexed for MEDLINE]).
- Yasukawa, K., M. Takido, T. Matsumoto, M. Takeuchi, and S. Nakagawa, 1991. Sterol and triterpene derivatives from plants inhibit the effects of a tumor promoter and sitosterol and betulinic acid inhibit formation in mouse skin two-stage Carcinogenesis. *Oncology*, 48: 72-76. (<http://www.ncbi.nlm.nih.gov/pubmed/1898988>), (PMID: 1898988 [PubMed - indexed for MEDLINE]).
- Youssef, S.A. and K.M. Ashry, 1999. Toxicopathologic evaluation of hepatoprotective effect of crude garlic and *Nigella sativa* seed extracts against aflatoxicosis in rats. *Alex. J. Vet. Sci.*, 15: 521 - 531. (ISSN: 1110-2047).